

Foundations of Medicine Sessions 164 & 167

Group Case



Case Studies Making connections between Genetics, Molecular Biology, & Biochemistry

With recent advances in the integration of various disciplines of molecular science and technological developments in genetic analysis, it is now possible to implement truly “personalized” medicine. The growing adoption of “Precision Medicine” involves the full understanding of a patient, including their own specific molecular pathology and disease etiology, which can help to establish an accurate diagnosis and to select an effective therapy.

NCBI has long had online resources for biologists to explore what is known about a biological molecule including its structure and function, but has recently developed clinically-focused resources enabling scientists and clinicians to integrate known molecular biological information with clinically-relevant genetic variations.

In Wednesday’s Session:

- We discussed the state of clinical practice with regard to the application of Precision Medicine principles (examining a patient’s specific molecular pathology).
- Together we explored a real-world case study and followed a workflow to discover the patients’ molecular pathology for an undiagnosed/misdiagnosed problem.

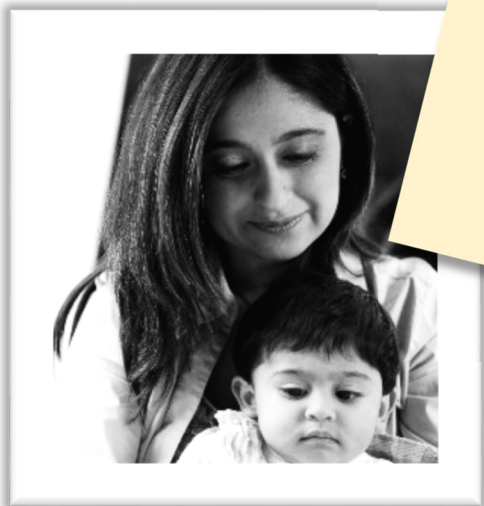
Before Friday’s Session:

- **There is a practice case study to solve!**

In Friday’s Session:

- You will work in groups to practice solving additional case studies as independent exercises – and we will discuss the cases and discover the underlying cause of pathology in these real patients.
- We’ll compare what is happening at the molecular level in other patients that have seemingly related cases.



Raven




*Here's the patient's referral
and the genetic test results
for the
molecular pathology work up.*

*Needs to be done and
ready for presentation
by Friday!*

Thanks

Patient Information	
Patient Name RAVEN	Patient Barcode Sticker 
DOB, Medical Record Number (MRN) <div style="background-color: #cccccc; width: 100px; height: 15px; display: inline-block;"></div> <div style="background-color: #cccccc; width: 100px; height: 15px; display: inline-block;"></div>	
Requesting Provider	
Assigned Provider/Practice Name: Jane Ferreira, MD / MyClinicalService	Specialty/Department: Family Practice
Address: 900 23rd St NW Washington, DC 20037	Phone: (202) 555-1212 Facsimile #: (202) 555-1212
Consultant Provider	
Provider's Name: to be assigned	Specialty/Department: Molecular Science/M1 Training
Address: 2300 I St NW, Suite 201 Washington, DC 20052	Phone: (202) 555-1212 Facsimile #: (202) 555-1212
Referral Information	
Authorization No:	Authorization Type:
Reason for Referral: Evaluation of Hemophilia Carrier Status	
Diagnosis: Z14.8 – Asymptomatic Carrier of “Other” Genetic Disorder	
<p>Clinical Notes: 25 year old female was referred for consultation after her son was diagnosed with Hemophilia. In addition to the identification of a specifically diagnosed proband, she mentioned a 6 year old brother who died of a “brain bleed” who she suspects might have “had Hemophilia too”. She described her own history as unremarkable, but upon questioning mentioned that she required a blood transfusion after “normal childbirth” and has “always had really, really heavy periods”. She requested evaluation of potential carrier status for family planning purposes.</p> <p>A blood sample has been sent out for analysis with a Hemophilia genetic testing panel. The genetic test result report will be faxed to the Molecular Science/M1 Training program for evaluation.</p> <p>Please consult with the patient and send a copy of the final report back to this office. Thanks.</p>	
Procedures: Variant Interpretation – Molecular Impact Characterization	
Visits Allowed: 3	
Unit Type: V (VISIT)	
Referral is Valid Until: 09/30/2018	
Notes: Patient must arrive 30 minutes early, with a picture ID, Insurance card and have a copy of this referral. Please bring a list of medications the patient is taking with you to this appointment (including over the counter).	
Please send the final report by Fax to: (202) 555-1212	
Signature: 	
Ferreira, Jane, MD on 08/29/2018 at 1:37 PM EDT	

Specimen Number	Specimen Type Peripheral Blood		Control Number	Account Number	Account Phone Number	Route	
Patient Last Name							
Patient First Name Raven		Patient Middle Name					
Patient SS#	Patient Phone	Total Volume					
Age (Y/M/D) 25 y.o.	Date of Birth	Sex Female					Fasting
Patient Address			Indication: Hemophilia B carrier Family History: Family history of uncontrolled bleeding Ethnicity: Native American, Navajo Tribe				
Date and Time Collected	Date Entered	Date and Time Reported		Physician Name Jane Ferreiro, MD	NPI	Physician ID	
Hemophilia Mutation Evaluation			Tests Ordered				
<p style="text-align: center;">General Comments</p> Please send a copy of the final report to the Molecular Science/M1 Training office via Fax at (202) 555-1212							

Clinical test results for DNA Hemophilia Mutation Evaluation

GENE	TEST RESULTS	EXPLANATION
F8 (Xq28)	Arg15 Arg15Ter Heterozygous Carrier Status	<p>This result confirms the status of carrier for Hemophilia A. This result should be interpreted in the context of clinical presentation and results of other laboratory tests (e.g., APTT, Factor VIII Activity, etc.).</p> <p>A PCR/sequencing study has detected one copy of the Arg15Ter (F8:g.5214C>T, c.43C>T or p.Arg15Ter) variation. The Arg15Ter variation is a C to T change at nucleotide position 5214 of the F8 gene and 43 of the F8 mRNA transcript. This forms a premature stop (termination) codon at amino acid position 15 resulting in an abnormally short or truncated protein.</p> <p>Female carriers of Hemophilia A have one normal X chromosome and one abnormal X chromosome. The normal X chromosome produces a certain amount of factor IX clotting factor. This protects carriers from the most severe form of hemophilia, however the levels of circulating clotting factors among carriers are very wide.</p>
F9 (Xq27.1)	Negative	

INDICATIONS FOR TESTING

Individuals with a diagnosis of hemophilia B, appropriate at-risk female relatives of probands with identified mutations, and hemophilia B carriers with genetic counseling, are candidates for testing.

METHODOLOGY

Factor IX sequencing: All coding exons (1-8) and associated intron junctions of the Factor IX gene are analyzed by direct DNA sequence analysis using an automated fluorescent sequencing machine. When a mutation is detected, confirmation is carried out on an independent amplification of PCR using a second prep (B-prep) by sequencing in the opposite direction. If no mutation is found, sequence analysis is performed in both directions.

PERFORMANCE

Factor IX sequencing: From previous experience, we have been able to detect factor IX gene mutations in about 99% of individuals with the diagnosis of hemophilia B with specificity of mutation detection in probands and carrier detection is also estimated to be greater than 99%.

LIMITATIONS

The sequence analysis will not detect mutations located in regions of the Factor IX gene that are not analyzed (non-coding exon regions, intron regions other than the splice junctions, and upstream and downstream regions). The sequencing method also will not detect gross genetic alterations including most duplications, inversions, or deletions (in females). Some sequence alterations that may be detected (such as those causing missense or synonymous changes) will be of unknown clinical significance.

CLINICAL DESCRIPTION

Hemophilia B is characterized by deficiency in factor IX clotting activity that results in prolonged oozing after injuries, tooth extractions, or surgery, and delayed or recurrent bleeding prior to complete wound healing. This is an X-linked recessive bleeding disorder with an incidence of about 1 per 30,000 live male births. Hemophilia B affects males, however, all male offspring will be normal, and although all female offspring will be obligatory carriers, they rarely have symptomatic bleeding. In contrast, female offspring of carriers of hemophilia B have a 50% chance of being carriers themselves, and each male offspring has a 50% chance of being affected.

The age of diagnosis and frequency of bleeding episodes are related to the level of factor IX clotting activity. In severe hemophilia B, spontaneous joint or deep-muscle bleeding is the most frequent symptom. Individuals with severe hemophilia B are usually diagnosed during the first two years of life; without prophylactic treatment, they may average up to two to five spontaneous bleeding episodes each month. Individuals with moderate hemophilia B seldom have spontaneous bleeding; however, they do have prolonged or delayed oozing after relatively minor trauma and are usually diagnosed before age five to six years; the frequency of bleeding episodes varies from once a month to once a year. Individuals with mild hemophilia B do not have spontaneous bleeding episodes; however, without pre- and post-operative treatment, abnormal bleeding occurs with surgery or tooth extractions; the frequency of bleeding may vary from once a year to once every ten years. Individuals with mild hemophilia B are often not diagnosed until later in life. In any individual with hemophilia B, bleeding episodes may be more frequent in childhood and adolescence than in adulthood. Approximately 10% of carrier females are at risk for bleeding (even if the affected family member has mild hemophilia B) and are thus symptomatic carriers, although symptoms are usually mild. After major trauma or invasive procedures, prolonged or excessive bleeding usually occurs, regardless of severity.

REFERENCES

4. Yoshitake S, Schach BG, Foster DC, et al: Nucleotide sequence of the gene for human factor IX (antihemophilic factor B). *Biochemistry* 1985 July 2;24(14):3736-3750
5. Giannelli F, Green PM, Sommer SS, et al: Haemophilia B: database of point mutations and short additions and deletions-eighth edition. *Nucleic Acids Res* 1998 Jan 1;26(1):265-268
6. Ketterling RP, Bottema CD, Phillips JA 3rd, et al: Evidence that descendants of three founders constitute about 25% of hemophilia B in the United States. *Genomics* 1991 Aug;10(4):1093-1096

Researching the Referral

1. To learn more about the preliminary diagnosis, **go to the NCBI website** (<https://www.ncbi.nlm.nih.gov> or “google” NCBI to find the homepage) and **search NCBI’s MedGen database with: Hemophilia [ExactTitle]**

In the “Term Hierarchy” section you can see more specific sub-types of “Hemophilia” -two major forms of hereditary disease are displayed. Click the names of the diseases to open the MedGen records to read about each hereditary sub-type.

WHAT IS/ARE THE MAJOR DIFFERENCES IN THE TWO SUB-TYPES OF HEREDITARY HEMOPHILIA?

WHICH ONE WAS SUSPECTED IN HER SON AND THEY WERE TESTING FOR IN RAVEN?

Understanding the Genetic Test Results

2. **WHAT IS THE SPECIFIC AFFECTED GENE AND VARIATION IDENTIFIED IN RAVEN?**
(Read the results, sometimes it is really helpful!)

THEY ASSESSED TWO COPIES OF THE GENE IN RAVEN, WHAT DID THEY FIND FOR EACH OF THOSE TWO COPIES?

WHAT DOES THE RESULTS OF THE GENETIC TEST MEAN FOR RAVEN?

You can find out what various genetic testing laboratories, clinical genetic organizations, and OMIM are claiming with regard to health-related impact for these genetic variations in the [ClinVar database](#).

You can search with a Gene Symbol and nucleotide or protein change, an rsID or an HGVS expression, for example type:

F8 Arg15Ter

Molecular Biology Research

INFORMATION ABOUT THIS GENE FROM HUMAN-CURATED SOURCES:

3. On the MedGen record, [click the link for the gene](#) identified as having a variant in Raven.
WHAT DOES THIS GENE NORMALLY DO?

4. From the Gene record, [scroll down to the General gene information>Gene Ontology section](#) to learn more about the protein produced from this gene. This section displays terms for where this gene product is likely to be found within a cell (Component), what processes it is often involved in (Process), and what it does (Function).

**WHAT TYPE(S) OF PROCESS(ES) IS/ARE THIS PROTEIN NORMALLY INVOLVED WITH?
DOES THIS MAKE SENSE BASED ON THE SUMMARY OF THE GENE THAT YOU JUST FOUND?**

**WHAT SPECIFIC FUNCTION(S) DOES THIS PROTEIN HAVE?
DOES THIS MAKE SENSE BASED ON THE SUMMARY OF THE GENE THAT YOU JUST FOUND?**

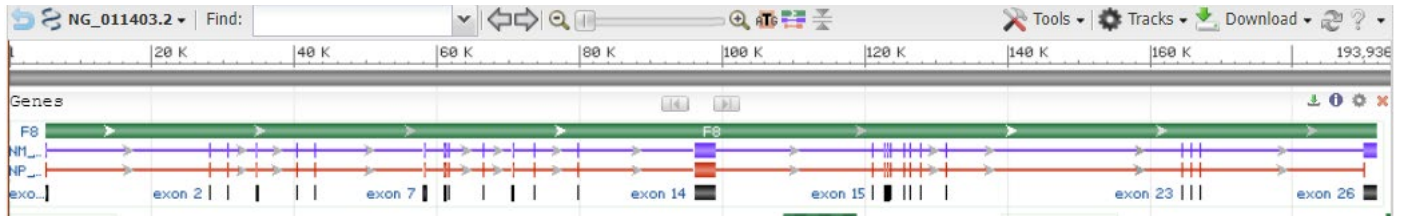
IN WHICH COMPONENT(S) (SUB-CELLULAR LOCATION) IS THIS PROTEIN NORMALLY FOUND?

5. Now find the [Expression section](#) to see in which tissues this gene is expressed.
IN WHICH TISSUES HAS THIS GENE BEEN FOUND TO BE EXPRESSED?

BASED ON WHAT YOU READ ABOVE, ABOUT THE FUNCTION AND PURPOSE OF THIS PROTEIN, WHAT WOULD YOU PREDICT TO FIND IN THE PROTEIN SEQUENCE? (HINT: if a protein is made in a cell type/tissue, but functions elsewhere....how does it get there? Ask Dr. Elliott if you *really* can't remember.)

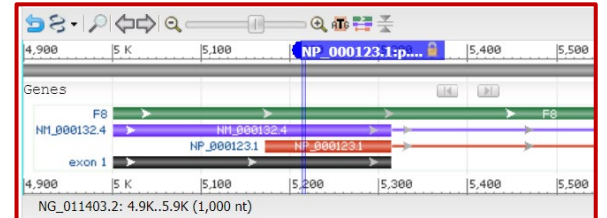
INFORMATION ABOUT THIS GENE DETERMINED FROM SEQUENCE-BASED SOURCES:

- From the Gene record, (on the right-hand side of the page) **click the “RefSeqGene” link** to see the “Graphic” view of the gene structure defined on the chromosome on a RefSeqGene nucleotide page.



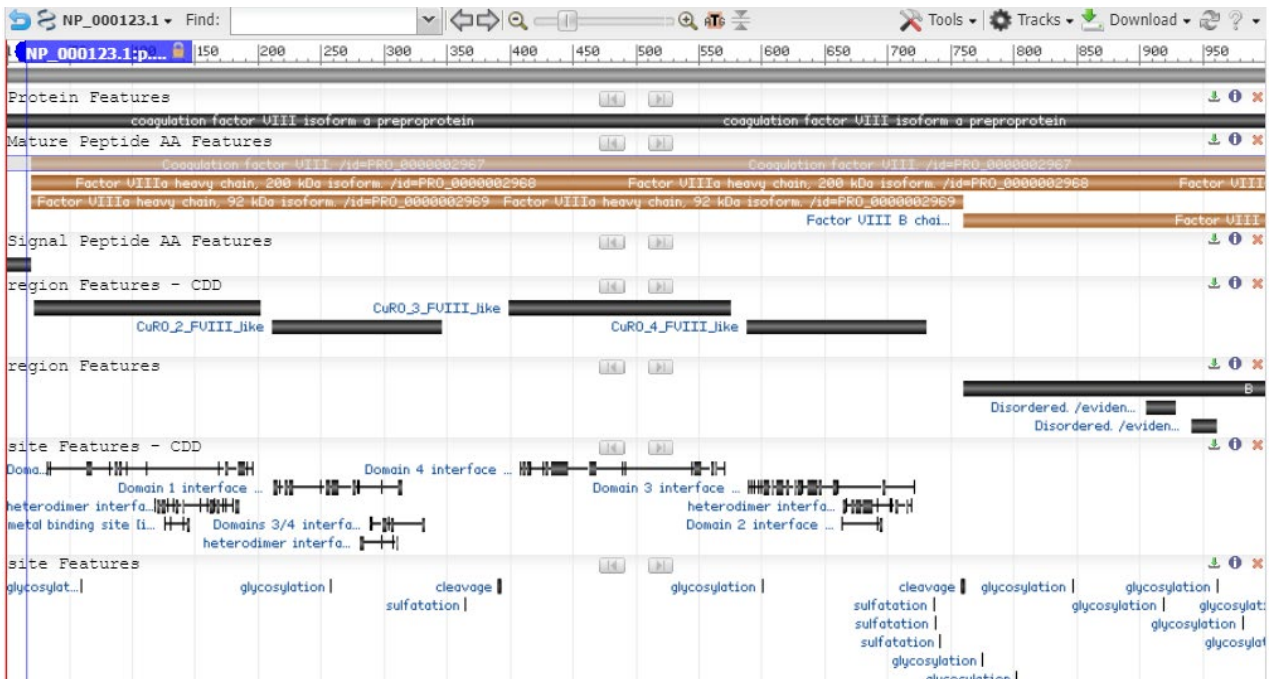
WHERE IS RAVEN’S GENETIC VARIANT GENE AND IN THE MRNA?

(On the picture above or on your screen – draw or visualize a vertical line at the variant’s position. You can type in the variant’s gene position, from the genetic test result, into the “Find” box to automatically zoom in!)



BASED ON THE POSITION OF THE VARIANT IN THE GENE, WHAT IS THE MOST LIKELY MECHANISM FOR IMPACTING THE FINAL GENE PRODUCT? *(alter gene expression, influence transcript processing, or change encoded protein sequence)*

- On the Gene page, (on the right-hand side) **click the “RefSeq Proteins” link** and then pick the full-length pre-processed version of the protein and **Click “Graphics”** to see a graphical view of the annotated regions curated on the protein sequence. The information shown in these “tracks” of this view can help you to learn more about this protein.



WHERE IN THE PROTEIN SEQUENCE IS RAVEN’S GENETIC VARIANT LOCATED?

*(On the picture shown above or on your screen – draw or visualize a vertical line at the position of each if the variants. You can learn more about the main functional regions of the protein **click “Identify Conserved Domains”**)*

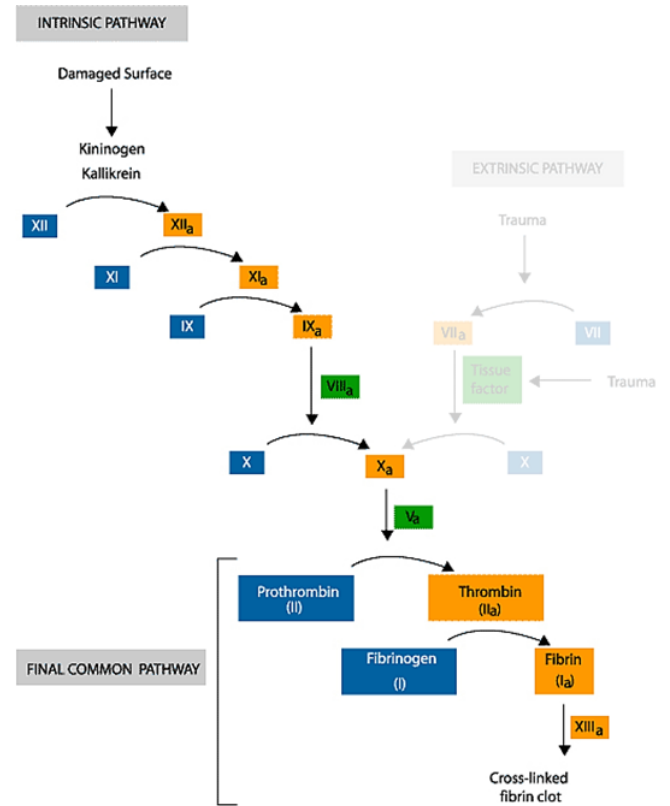
WHAT MIGHT BE THE IMPACT OF THE GENETIC VARIATION ON THE PROTEIN’S FUNCTION?

8. You found out earlier that this protein is not only involved, but critical for inducing the formation of a clot to stop bleeding.

Here is a graphic of the Clotting Cascade pathway →

HOW DO YOU THINK THE PROTEIN PRODUCED BY RAVEN'S GENE WITH THE VARIANT ULTIMATELY IMPACTS HER CLOTTING CASCADE?

Unlike her son, who has had pretty severe bleeding problems, **WHY DO YOU THINK RAVEN HASN'T REALLY HAD TOO MANY ISSUES, DESPITE HAVING A GENE CONTAINING A PATHOGENIC VARIANT?**



(Can you convert these Roman Numerals into Arabic Numerals to figure out which step is affected?)

She originally requested a genetic test for “Family Planning” purposes. **WHAT ARE THE CHANCES OF HAVING ANOTHER SON WITH HEMOPHILIA A?**

Finally, although she has been designated as a “Carrier” and has not been diagnosed with the disease, **IS THERE A REASON FOR RAVEN TO KEEP IN MIND THAT SHE DOES HAVE ONE COPY WITH A PATHOGENIC VARIANT? UNDER WHAT CIRCUMSTANCES COULD THIS BE IMPORTANT TO KNOW?**

SUMMARY QUESTIONS – You should be prepared to discuss these specific questions.

Introduce your patient to the class!



*Who is she? What is her story?
(see the referral form)*

What was the preliminary diagnosis and the rationale for it?

(see the referral form & NCBI's MedGen database)

What did the genetic test find and how does this relate to the preliminary diagnosis?

(see the genetic test result form & NCBI's ClinVar database)

What is the implicated/affected gene and what is its normal function?

(NCBI's Gene database should help!)

Where in the gene and gene product is the patient's genetic variant located?

(Where in the gene? In what part of the mRNA? Where in the protein? In what functional part of the protein?)

What is the molecular impact of the genetic variant on the gene product?

(What do you think the variant ended up doing to the protein structurally?)

What do you think might be the functional impact of the variant on the gene product and in the patient?

(What impact do you think the variant had on the function of the protein? How might this relate to the patient's symptoms?)

Now that you're done.....SELF-ASSESSMENT TIME!

My initial ideas about this case:

*(Why did I think this?
How confident was I?)*

What did I miss?

*(Why did I miss it?
How could I have thought about it differently?)*

What specific content areas do I need to review?